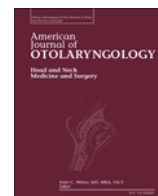




Contents lists available at ScienceDirect

American Journal of Otolaryngology–Head and Neck Medicine and Surgery

journal homepage: www.elsevier.com/locate/amjoto

Intestinal permeability and Ménière's disease

F. Di Bernardino^{a,*}, D. Zanetti^a, E. Ciusani^d, C. Caccia^d, V. Leoni^d, U. De Grazia^d, E. Filippini^b, L. Elli^c^a Audiology Unit, Head and Neck Department, Fondazione IRCCS Ca' Granda Ospedale Maggiore Policlinico, Department of Clinical Sciences and Community Health, University of Milan, Italy^b U.O.C. Direzione Professioni Sanitarie, Fondazione IRCCS Ca' Granda Ospedale Maggiore Policlinico, Department of Clinical Sciences and Community Health, University of Milan, Italy^c Center for Prevention and Diagnosis of Celiac Disease, Fondazione IRCCS Ca' Granda Ospedale Maggiore Policlinico, Milan, Italy^d U.O. Laboratorio di Patologia Clinica e Genetica Medica, Neurology and Neuroscience R17 Fondazione IRCCS Istituto Neurologico Carlo Besta, Milan, Italy

ARTICLE INFO

Article history:

Received 8 October 2017

Available online xxxx

Keywords:

Ménière

Double sugar test

Fecal calprotectin

Intestinal permeability

Lactulose

Mannitol

ABSTRACT

Purpose: Ménière disease (MD) is a multifactorial chronic disabling condition characterized by episodic vertigo, ear fullness, and hearing loss. MD patients often complain of aspecific gastrointestinal symptoms associated with autonomic dysregulation, frequently outweighed by the otological manifestations. Dietary modifications have been reported to improve the typical MD symptoms in some cases. Our purpose was to test the urinary levels of lactulose and mannitol (double sugar test) and the fecal calprotectin, both markers of altered intestinal permeability, in subjects with definite MD in an active and inactive stage.

Materials and methods: Twenty-six with definite unilateral MD were studied: 14 patients were symptomatic for at least 3 months with moderate to severe vertigo spells and a functional level ≥ 4 ; 12 patients had been asymptomatic (no vertigo spells) for at least 3 months and had a functional level = 1 at the time of testing. Twenty healthy volunteers were recruited as “control group”.

Results: Lactulose and mannitol absorption was significantly increased in the symptomatic MD patients compared to the asymptomatic group ($p < 0.02$ and $p < 0.004$, respectively) and to the controls. FC were also higher than normal only in the symptomatic group. ($p < 0.01$).

Conclusions: An altered intestinal permeability, according to the two assays, was found only in symptomatic MD patients. The rationale for a possible relationship between MD and intestinal permeability is forwarded. The double-sugar test and FC quantification might be implemented in the MD diagnostic workup.

© 2017 Elsevier Inc. All rights reserved.

1. Introduction

Ménière disease (MD) is a multifactorial chronic disabling condition characterized by episodic vertigo, ear fullness, and hearing loss. The underlying pathologic condition has been identified in the endolymphatic hydrops, but its pathophysiology is not yet fully understood, as a causative role in the disease has been ascribed to many different etiological factors.

Several authors reported a significant improvement of the vestibular symptoms by generic or specific dietary modifications in MD patients [1,2,3]. A possible link between food allergies and MD symptoms has been firstly hypothesized in 1923; [4] after, that, since the 70's, targeted elimination diets for documented food allergies were proposed [5] after corroborating proof that avoidance might prevent an immune-mediated reaction to the inner ear [6]. However, double blinded food challenge

tests [7] are very difficult to be implemented because of long follow-up required and the highly unpredictable time course of the symptoms in MD patients [8,9].

On the other hand, most MD patients complain of aspecific gastrointestinal symptoms, such as diarrhea, abdominal pain, dyspepsia, and weight fluctuation; [10] the association of MD with gastrointestinal lesions has also been described many years ago [11]. Given the enhancement of gastroenteric mucosal permeability during episodes of local inflammation, and the frequency of reported gastrointestinal symptoms in MD patients, we designed to evaluate the intestinal permeability by means of a couple of simple assays: a) the “double sugar test” is a well-known validated not-invasive method; it is easily applicable, independently from the etiology; [12] b) the dosage of fecal calprotectin (FC) in the stools has been recently proposed as a marker for differentiation between functional and inflammatory gastrointestinal disorders [13,14]. FC is a calcium binding protein of the S100 family, found mainly in neutrophils but also in other white blood cells [15,16].

The aim of this study was to evaluate “double sugar test” and FC in two groups of definite MD patients in the active versus quiescent phase of the disease and compare them to those obtained in normal controls.

* Corresponding author at: Audiology Unit, Dept. of Clinical Sciences and Community Health, Fondazione IRCCS Ca' Granda Ospedale Maggiore Policlinico, Università degli Studi di Milano, Via Pace 9, 20122 Milan, Italy.

E-mail address: federica.diberardino@unimi.it (F. Di Bernardino).

2. Material and methods

2.1. Patients

The study included 26 consecutive unilateral definite MD patients, recruited at the outpatient clinic of the Vestibular Disorders Unit in a tertiary referral university Hospital. They fulfilled all criteria for definite MD according to the consensus paper of the international societies revision [17] of the AAO-HNS guidelines; [18] all of them were in the clinical subgroup 1, stage 2 or 3 [19]. Patients were negative for retrocochlear lesion at magnetic resonance imaging. Those who had previously received systemic steroids or intratympanic injections (either gentamicin or steroids) were excluded from the study, as well as those following specific diets or being treated with steroids, gastrointestinal drugs and/or antihistamines. A positive history of gastrointestinal disease or familiarity for celiac disease or intestinal bowel disease, of abnormal thyroid hormones, of malignant tumours or autoimmune diseases was exclusion criteria. Hypertension was not an exclusion criterium, if well controlled by drugs.

Fourteen patients (6 females, 8 males, mean age: 45.3 ± 11.2 years) were defined as “symptomatic” if they had at least two major episodes of vertigo per month with sensorineural hearing loss and aural fullness during the 3 months prior to the admission and a functional level (FL) ≥ 4 , according to the consensus paper guidelines [18]. Twelve patients (4 females, 8 males, mean age: 44.0 ± 14.4 years) were considered “asymptomatic” if they had been free from symptoms for at least 3 months at the enrolment time and a FL of 1.

Twenty healthy volunteers (10 females, 10 males, mean age: 39.0 ± 16.4 years) served as a “control group”, in order to check the reference values. All healthy subjects were normally hearing, reported a negative familiar and personal history of vertigo or dizziness, and never suffered from otological diseases.

The participation in the study was voluntary and all the subjects were not paid for it. The work was carried out in accordance with the Helsinki Declaration of 1975, as revised in 2013, including, but not limited to, the absence of potential harm to participants, their guaranteed anonymity, and written informed consent.

2.1.1. Double sugar test

The intestinal permeability was assessed using a double sugar test. In principle, the test is based on the measurement of the urinary excretion of orally administered non-metabolized sugar probe molecules [20]. The urinary content of lactulose and mannitol can be considered an indirect index of the intestinal absorption [21]. When awakening in the morning, each patient was instructed to collect a pre-test urine sample. Then, they drank a solution containing 5 g lactulose plus 5 g mannitol dissolved in 100 ml water. Urines were collected during the next 5 h and kept refrigerated at $2-7$ °C. Patients were instructed to avoid eating (not even a chewing-gum), drink or smoke during the test, but were allowed to drink a fixed dose of water (200 ml), only after 2 h. Total urine volume was recorded on completion of the test and a 10 ml fraction was stored at -20 °C until analysis.

Results were expressed as the percentage of recovery of the ingested dose of sugars.

2.1.2. Laboratory double sugar test analysis

Mannitol and lactulose concentrations were measured by gas chromatography mass spectrometry (GC-MS) [22]. A small aliquot (100 μ l) of each urine sample was added with 200 μ l of acetonitrile plus internal standards [methyl alfa-D-mannopyranoside 100 μ l (0.5 μ g/ μ l); D-(+)-turanose 50 μ l (0.1 μ g/ μ l)] in a glass tube. After evaporation under a gentle stream of nitrogen, the mixture was oximated by adding 100 μ l of a 25 μ g/ μ l solution of hydroxylamine in anhydrous pyridine and heated to 70 °C for 30 min. Sugar oximes were derivatized with 100 μ l of Chlorotrimethylsilane, N,O-bis (trimethylsilyl) trifluoroacetamide BSTFA with 1% Trimethylchlorosilane, Trimethylsilyl chloride TMCS at

70 °C for 30 min. Silylated derivatives, sealed in autosampler vials, were analyzed by a Clarus 600 GC-MS system equipped with an Elite column (30 m \times 0.32 mm id \times 0.25 mm film; Perkin Elmer, USA). Injection was performed with 1:20 split using helium (1 ml/min) as carrier gas. The temperature program was as follows: an initial temperature of 150 °C held for 2 min, followed by a linear ramp of 10 °C/min to 300 °C held for 5 min. The mass spectrometer operated in full mass scan. Peak integration was carried out manually and metabolites quantified against internal standards using calibration curves for the listed sugars. Based on preliminary studies, the assay was linear up to 500 μ g for mannitol and 50 μ g for lactulose. Intra-assay CV% was $\sim 3\%$ for the two biomarkers and recovery up to 99%.

2.1.3. Fecal calprotectin

In normal subjects, FC values < 40 μ g/g are considered as negative; values between 40 and 60 μ g/g as borderline and values > 60 μ g/g as positive. A cut-off of 50 μ g/g has a sensitivity and a specificity $> 80\%$, and a positive and negative predictive values of 70 and 90%, respectively [23]. Values between 40 and 60 μ g/g suggest a possible intestinal inflammatory process; values ≥ 60 μ g/g indicate a significant inflammatory process of the intestinal mucosa.

2.1.4. Laboratory FC analysis

Standardised laboratory protocol (ELISA) for FC analysis was used. A 100 mg quantity of stool was weighed and dispensed into an analysis pot using a 10 μ l inoculation loop. As per local protocol, first morning sample was requested. The exact weight was recorded and 5 ml of extraction buffer was added. The samples were then vortexed for 30 min in order to ensure complete dissolution, and then centrifuged using an Eppendorf centrifuge. The supernatant was then removed for analysis using the IDK® Calprotectin ELISA method (Immunodiagnostik, Bensheim, Germany).

2.1.5. Statistical analysis

Statistical analysis was performed using the SPSS statistical package version 24.00 (SPSS Inc., Chicago, Illinois). The significance of difference between the two groups compared with each other and with controls was evaluated by student *t*-test for independent samples. Results are expressed as means and standard deviations (SD), median and first and third quartiles. A $p < 0.05$ was considered statistically significant. Spearman correlation test was performed to investigate the association between the variables.

3. Results

The two MD groups resulted homogeneous for age, sex, stage, onset time of MD, ear side, degree of hearing loss, absence of comorbidities; they differed only for the number of vertigo episodes during the last 3 months and the functional level, accordingly to the inclusion/exclusion criteria.

A significant increase of both sugars was found in symptomatic MD compared to the asymptomatic MD patients and to the controls, with a most consistent rise observed for mannitol in the symptomatic MD group. ($p = 0.004$) (Fig. 1).

Similarly, the levels of FC were significantly higher in the symptomatic MD group resulting in a range between 46 and 380 μ g/g (median: 120.4 μ g/g) compared to a range of 2–80 μ g/g (median: 28.5 μ g/g) in the asymptomatic MD subjects ($p = 0.01$); all subjects in the control group showed normal values (< 40 μ g/g). (Fig. 2) A positive correlation was observed between FC levels and mannitol absorption values. ($r = 0.768$).

Table 1 summarizes the results of the double sugar test and FC. The overall outcomes were comparable between asymptomatic MD and the control groups, except for the amount of urinary lactulose that resulted slightly but significantly higher in the former than in controls (median: 1.2% [range, 0.0–1.8] vs 0.5% [0.13–1.1]). ($p = 0.02$).

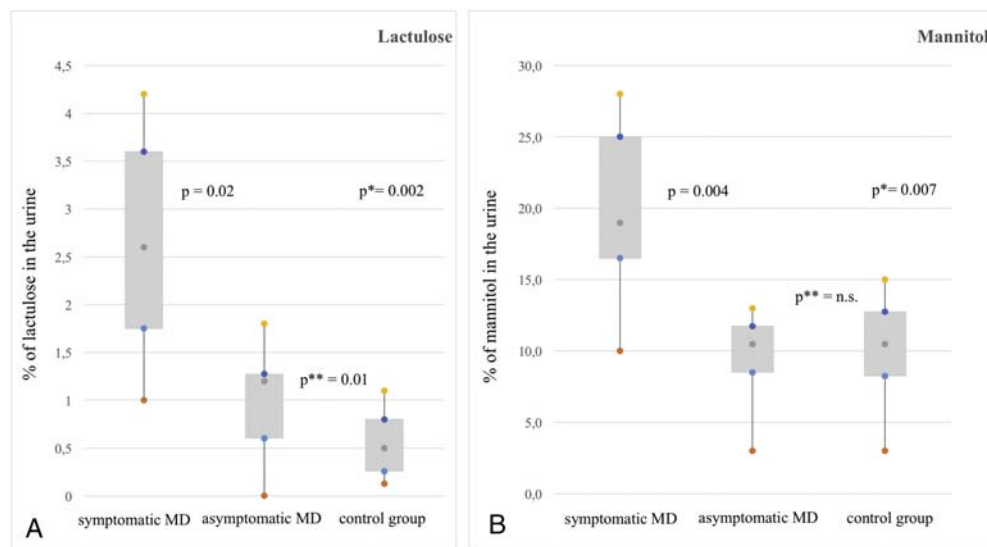


Fig. 1. The boxplots represent the urinary levels of lactulose and mannitol in the three groups.

4. Discussion

Several conditions often act as comorbidities of MD, including arthritis, psoriasis, gastroesophageal reflux disease, irritable bowel syndrome, and migraine [17]. On the other hand, scanty and heterogenous reports have included dizziness, [24] numbness [25,26] and/or sensorineural hearing loss [27,28,29] among the various comorbid symptoms in irritable bowel syndrome and/or intestinal bowel disease [30]. Since 1997, Derebery [31] supposed that there was considerable clinical and immunological evidence in favour of a probable role of allergies in MD symptoms: inhalant and food allergies have both been linked with the symptoms. In a previous work, we found an increased rate of atopic subjects among the MD patients (82.7%): food most frequently inducing hypersensitivity in MD were wheat, cow's milk protein and tomato [32].

Dietary modifications have sometimes proven effective in improving the typical MD symptoms [1]. More recently, Heather stated that allergy avoidance and immunotherapy should be considered as part of the treatment plan to help controlling MD symptoms [33].

As food intolerance is frequently expressed in atopic patients, our hypothesis in the present study was that the acute MD crises might be associated with conditions of acutely altered intestinal permeability [2,5]. However, a proof of this correlation is difficult to assess, because MD patients often show hypersensitivity to many foods, and a combination of dietary deprivation and challenge tests require very long observation periods and repeated trials.

Our preliminary data seem to support the hypothesis that an impaired intestinal barrier function may be pathologically related with the active/inactive stage of MD symptoms.

In healthy subjects, <1% of lactulose passes the intestinal mucosal barrier, only through the intercellular junctions (a para-cellular pathway); practically, all lactulose remains unabsorbed and can be found in the stools. Conversely, under normal conditions, about 14% of mannitol is absorbed through the hydrophilic pores of the enterocytes. Diseases causing a definite damage to the villi (e.g. celiac sprue, giardiasis, etc.) determine a reduced absorption of mannitol whereas other chronic diseases of the bowel (e.g. Crohn's disease, ulcerative colitis) show an increased permeability to lactulose [12].

In both our symptomatic and asymptomatic MD patients, lactulose absorption was significantly increased compared to the control group. Interestingly, instead of a reduction, an increase of mannitol intake was observed in the symptomatic MD group. Similarly, also FC was higher in the symptomatic MD group. FC is a calcium-binding protein, classified as damage-associated molecular pattern protein that has

antimicrobial properties [34]. It is released mainly from neutrophilic leukocytes and secreted into the whole enteric tract during inflammation of the intestinal mucosa [15,16]. Therefore, it is a marker of inflammation of the intestinal tract. It is an expression of the activation of the mucosal innate defense system against a pro-inflammatory challenge, such as in irritable bowel syndrome patients. Interestingly, it can be detected also in the absence of macroscopic signs of inflammation [35].

The increased absorption of lactulose and mannitol, together with higher values of FC, clearly indicates an altered intestinal permeability in the active phases of MD. We might speculate that the impaired intestinal barrier function in symptomatic MD, could be possibly linked to a decreased tightness of the intercellular junctions caused by the inflammation, rather than by a direct damage to the intestinal cells. However, to confirm this hypothesis a further study with a longer follow-up in the same patients has been planned, in order to crosscheck the two groups.

A limit of this study is the small size of the sample group. Nevertheless, we are not aware of other studies addressing the possible link

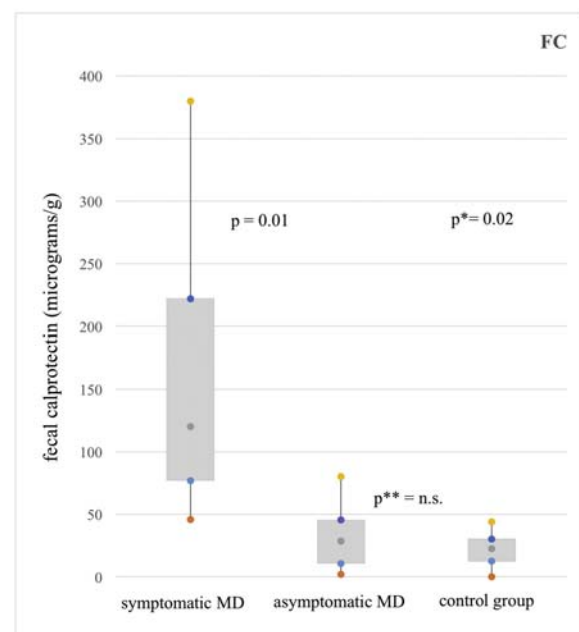


Fig. 2. The boxplots show the values of FC.

Table 1

Recovery ratio of the administered sugars (percentage of the ingested dose of sugars) and the amount of fecal calprotectin (FC).

Patients	Lactulose % (n.v. <1)	Mannitol % (n.v. <14)	FC µg/g (n.v. <40)
Symptomatic MD group	2.64 ± 1.3	20 ± 6.4	163.4 ± 119.4
Asymptomatic MD group	1.22 ± 0.34	9.5 ± 3.6	32.5 ± 27
Non-MD control group	0.56 ± 0.39	10 ± 4.3	21.8 ± 15.6

Mean ± standard deviation; normal reference values = n.v.

between altered intestinal permeability and symptomatic MD and we will proceed with the recruitment of new patients in order to increase the statistical power of the research. These interesting results might also allow to think that testing these parameters after specific therapies or in the intercritical phase of MD in the same subjects may provide interesting insights in the follow-up. We also advocate further trials by other centres in order to assess if the investigation of intestinal permeability and FC can be added to the current clinical practice, aimed at identifying those MD patients who would benefit from deprivation diets.

References

- [1] Luxford E, Berliner KI, Lee J, et al. Dietary modification as adjunct treatment in Ménière's disease: patient willingness and ability to comply. *Otol Neurotol* 2013; 34(8):1438–43.
- [2] Wu H, Gao Z. Vertigo with dysautonomia and serious allergy: an unusual case of juvenile Ménière's disease. *Int J Pediatr Otorhinolaryngol* 2015;79(12):2438–41.
- [3] Di Berardino F, Filippini E, Alpini D, et al. Ménière disease and gluten sensitivity: recovery after a gluten-free diet. *Am J Otolaryngol* 2013;34(4):355–6.
- [4] Duke W. Meniere's syndrome caused by allergy. *JAMA* 1923;81:2179–82.
- [5] Bryan WT, Bryan MP. Clinical examples of resolution of some idiopathic and other chronic disease by careful allergic management. *Laryngoscope* 1972;82:1231–8.
- [6] Derebery JM, Berliner KI. Allergy and its relation to Meniere's disease. *Otolaryngol Clin North Am* 2010;43:1047–58.
- [7] Elli L, Villalta D, Roncoroni L, Barisani D, Ferrero S, Pellegrini N, Bardella MT, Valiante F, Tomba C, Carroccio A, Bellini M, Soncini M, Cannizzaro R, Leandro G. Nomenclature and diagnosis of gluten-related disorders: A position statement by the Italian Association of Hospital Gastroenterologists and Endoscopists (AIGO). *Dig Liver Dis* 2017;49(2):138–46.
- [8] Smecuol E, Vazquez H, Sugai E, et al. Sugar tests detect celiac disease among first-degree relatives. *Am J Gastroenterol* 1999;94(12):3547–52.
- [9] Duerksen DR, Wilhelm-Boyles C, Parry DM. Intestinal permeability in long-term follow-up of patients with celiac disease on a gluten-free diet. *Dig Dis Sci* 2005;50(4):785–90.
- [10] Crane AW. Extra-abdominal affections giving gastro-intestinal symptoms, with special reference to the Meniere syndrome. *Radiology* 1928;11:447–52.
- [11] Wyburn-Mason R. Association of Gastro-duodenal lesions with Ménière's syndrome. *Br Med J* 1959;1(5114):79–83.
- [12] Sequeira IR, Lentle RG, Kruger MC, et al. Standardising the lactulose mannitol test of gut permeability to minimise error and promote comparability. *PLoS ONE* 2014; 9(6):e99256.
- [13] Pieczarkowski S, Kowalska-Duplaga K, Kwinta P, et al. Diagnostic value of fecal calprotectin (S100 A8/A9) test in children with chronic abdominal pain. *Gastroenterol Res Pract* 2016;2016:8089217 (Epub2016 Nov 15).
- [14] Waugh N, Cummins E, Royle P, et al. Faecal calprotectin testing for differentiating amongst inflammatory and non-inflammatory bowel diseases: systematic review and economic evaluation. *Health Technol Assess* 2013 Nov;17(55):xv–xix 1–211 <https://doi.org/10.3310/hta17550>. (Review).
- [15] Røseth AG, Fagerhol MK, Aadland E, et al. Assessment of the neutrophil dominating protein calprotectin in feces. A methodologic study. *Scand J Gastroenterol* 1992;27:793–8.
- [16] Aomatsu T, Yoden A, Matsumoto K, et al. Fecal calprotectin is a useful marker for disease activity in pediatric patients with inflammatory bowel disease. *Dig Dis Sci* 2011;56:2372–7.
- [17] Lopez-Escamez JA, Carey J, Chung WH, et al. Classification Committee of the Barany Society, Japan Society for Equilibrium Research; European Academy of Otolology and Neurology (EAONO), Equilibrium Committee of the American Academy of Otolaryngology-Head and Neck Surgery (AAO-HNS), Korean Balance Society. Diagnostic criteria for Ménière's disease. *J Vestib Res* 2015;25(1):1–7.
- [18] Committee on Hearing and Equilibrium guidelines for the diagnosis and evaluation of therapy in Ménière's disease, American Academy of Otolaryngology-Head and Neck Foundation, Inc. *Otolaryngol Head Neck Surg* 1995;113(3):181–5.
- [19] Frejo L, Martin-Sanz E, Teggi R, et al. Meniere's disease consortium (MeDiC). Extended phenotype and clinical subgroups in unilateral Meniere disease: a cross-sectional study with cluster analysis. *Clin Otolaryngol* 2017 Feb 6. <https://doi.org/10.1111/coa.12844>.
- [20] Bjarnason I, Macpherson A, Hollander D. Intestinal permeability: an overview. *Gastroenterology* 1995;108:1566–81.
- [21] Paganelli R, Fagiolo U, Cancian M, et al. Intestinal permeability in patients with chronic urticaria-angioedema with and without arthralgia. *Ann Allergy* 1991;66:181–4.
- [22] Farhadi A, Keshavarzian A, Holmes EW, et al. Gas chromatographic method for detection of urinary sucralose: application to the assessment of intestinal permeability. *J Chromatogr B Analyt Technol Biomed Life Sci* 2003;784(1):145–54.
- [23] National Institute for Health and Clinical Excellence. Fecal calprotectin diagnostic tests for inflammatory diseases of the bowel. NICE diagnostic guidance (DG11); 2013.
- [24] Igarashi M, MacRae D, O-Uchi T, et al. Cochleo-saccular degeneration in one of three sisters with hereditary deafness, absent gastric motility, small bowel diverticulitis and progressive sensory neuropathy. *ORL J Otorhinolaryngol Relat Spec* 1981; 43(1):4–16.
- [25] Catassi C. Gluten Sensitivity. *Ann Nutr Metab* 2015;67:16–26. <https://doi.org/10.1159/000440990>.
- [26] Volta U, Bardella MT, Calabrò A, et al. An Italian prospective multicenter survey on patients suspected of having non-celiac gluten sensitivity. *BMC Med* 2014;12:85.
- [27] Crawley SW, Shifrin Jr DA, Grega-Larson NE, et al. Intestinal brush border assembly driven by protocadherin-based intermicrovillar adhesion. *Cell* 2014;157(2):433–46.
- [28] Scharl M, Frei P, Fried M, et al. Association between Cogan's syndrome and inflammatory bowel disease: a case series. *J Crohns Colitis* 2011;5(1):64–8.
- [29] Froehlich F, Fried M, Gonvers JJ, et al. Association of Crohn's disease and Cogan's syndrome. *Dig Dis Sci* 1994;39(5):1134–7.
- [30] Gloddek B, Alexiou C, Ott R, et al. Inner ear disorders in inflammatory bowel disease – another extraintestinal complication? *Gastroenterology* 2000;118:4.
- [31] Derebery MJ. Allergic management of Meniere's disease: an outcome study. *Otolaryngol Head Neck Surg* 2000;122:174–82.
- [32] Di Berardino F, Cesarani A. Gluten sensitivity in Meniere's disease? *Laryngoscope* 2012;122:700–2.
- [33] Heather MW, Yuri A. The link between allergy and Ménière's disease. *Curr Opin Otolaryngol Head Neck Surg* 2014;22:227–30.
- [34] Nakashige T, Zhang B, Krebs C, et al. Human calprotectin is an iron-sequestering host-defense protein. *Nat Chem Biol* 2015;11:765–71.
- [35] Langhorst J, Junge A, Rueffer A, et al. Elevated human beta-defensin-2 levels indicate an activation of the innate immune system in patients with irritable bowel syndrome. *Am J Gastroenterol* 2009;104:404–10.